Applying pain theory in fish spine envenomation

David Muirhead

Key words

Envenomation, first aid, injuries, toxins, pain, marine animals

Abstract

Personal experience of catfish spine envenomation leads the author to question the long-accepted heat-labile toxin denaturation hypothesis as explanation for the established and very effective first aid treatment using hot water immersion of the envenomed limb. An alternative hypothesis compatible with contemporary pain theory is proposed.

Pain hypotheses in current usage, including Gate Control theory and Diffuse Noxious Inhibitory Control (DNIC) theory, have evolved substantially from observations that interference stimuli such as vibration, heat or cold, applied to the peripheral skin can induce pain relief at remote anatomical sites.

Have we overlooked the obvious in continuing to accept the hypothesis, entrenched in the diving medical community, 1,2,3 that heat-labile properties of fish spine toxins explain the well-documented analgesic effectiveness of hot water limb immersion in fish spine envenomation? A literature search has revealed a remarkable paucity of papers addressing this issue. Those that do, appear to assume that the proven heat lability of the few fish toxins so far analysed is the actual mechanism.

In April 2000, the author received a minor envenomation by an Estuary Catfish, *Cnidoglanis macrocephalus*, while snorkelling in an estuary south of Adelaide as described above.

As a South Australian coastal general practitioner, with occasional experience of treating mostly minor marine fish-spine injuries, I am familiar with the core first aid management using hot water (approx 46°C) immersion of the affected limb. I expected excellent pain relief as I placed my envenomed right forearm into a bucket of hot water,

after first testing the water with my contralateral hand to avoid burns.

My confidence was vindicated, with almost instantaneous pain relief. But I was puzzled as to why, if the hot water was indeed inactivating the toxin, the pain would recur so promptly and at the same intensity upon removal of my arm from the hot water. I initially reasoned that until all the venom had been denatured, pain would continue, but this begged the question: why the dramatic pain relief within seconds of immersion in the first place? Perhaps the toxin is reversibly inactivated by heat? It might be capable of reconstitution with falling temperature, at least until persistent exposure to heat effects a more permanent decomposition, for example by allowing irreversible binding of component molecules to tissue substrate.

This explanation fails to address the fact that whilst both my puncture wound and those of patients I'd treated by hot water immersion were small in external appearance, there could be little doubt that the sting had penetrated to a depth of at least some millimetres into the soft tissues. Diving medical texts recommend that water used for pain relief be in the 45–50°C range, yet human tissues other than perhaps the dermis, necrose before reaching these temperatures.

It seems improbable that exposure of the cutaneous portion of a puncture wound to such temperature would be capable of raising more deeply embedded subcutaneous or intramuscular residuae to temperatures sufficient to inactivate toxin without also causing significant tissue necrosis to the full depth of the puncture wound. This would in itself be very painful and thus defeat the objective. Further, all the fish spine wounds I've successfully treated by this method have been accompanied by sufficient localised oedema and serosanguinous ooze to make it unlikely in the first place that hot water could traverse the length of the puncture track, again discrediting this theory.

If one reviews current pain theory,^{4.5} support for the role of Gate Control and DNIC pain theories in fish spine envenomation may be found in the extensive range of fish species whose spine envenomations are known to respond to hot water immersion. This is specifically with regard to local pain relief as opposed to any systemic sequelae of envenomation. It even appears probable that envenomations from creatures in other phyla such as Cnidaria ⁶ will respond to thermal treatment, and application of icepacks is already an accepted first aid analgesic measure for jellyfish stings.

Examples do exist in the marine environment of identical or nearly identical toxins being utilized defensively by phylogenetically disparate organisms, notably tetrodotoxin in puffer fishes (vertebrates) and blue-ring octopi (invertebrates). However, it seems improbable that hundreds, even thousands of marine animal species share toxins so similar that they are all inactivated by such a conveniently small elevation in temperature.

A comprehensive worldwide literature search via Medline dating back to 1966, has failed to find a single study whose specific aim was to demonstrate heat-labile properties of non-scorpaeniform fish spine venoms. A limited number of papers were found purporting to delineate haemolytic, dermonecrotic, oedema-promoting, vasospastic, and lethal components of catfish venom and skin toxins.⁷

These include studies of the oriental catfish (Plotosus lineatus),⁸ North American species,^{9,10} and the comparative toxicity of two catfish genera Ictalurus and Schilbeodes.¹¹ None of the above studies addressed heat-lability.

As long ago as 1966, Pacy in his review of Australian catfish injuries proposed, largely on the basis of a single case report involving a long-tailed catfish (Family Plotosidae, genus Plotosus), that the venom had vasospastic properties as well as possibly transient neurotoxic effects.¹²

Pacy stated:

"Fish venoms tend to become rapidly inactive by change of pH (Wiener, 1960) and therefore instantaneous irrigation of the wound channel with sodium bicarbonate solution...is likely to destroy much of the poison."

and:

"As stingray venom is destroyed by temperatures above 60°C the possibility of this applying to catfish venom cannot readily be excluded. However, it is the great symptomatic relief that indicates hot bathing."

Pacy refers to earlier work specifically on stingray venom, and the quoted minimum temperature of 60°C needed to destroy venom would seem to negate the relevance of heatlability as the major reason for the effectiveness of hot water immersion in stingray envenomation.^{13,14}

However, Pacy's case report does contain the following statement:

"The patient repeatedly tried to take her hand out of the hot water, in order to get some sleep, but immediately the hand was withdrawn, the pain returned within a few seconds, disappearing only after renewed immersion."

This account perfectly matches my own experience, and is supported by a prospective observational case series of 22 fish stings, at least eight of which were from catfish, where hot water immersion treatment was completely effective in 73% of cases.¹⁵

The Estuary Catfish (*Cnidoglanis macrocephalus*) is the only marine member of the Plotosidae family known to occur in southern Australia, but most northern Australian catfish-spine injuries are also due to this family.¹⁶

The 'Poisindex Managements' 17 first aid treatment guidelines for catfish state:

"HOT WATER – The injured part should then be submerged in hot water at as high a temperature as the patient can tolerate without injury (less than 113 degrees F or 45 degrees C), for 30 to 90 minutes or more."

However, none of the three references provided contain proof of heat-lability of catfish venom. Indeed, Sutherland and Tibballs¹⁸ in their chapter titled *Venomous fish other than stonefish*, state:

"Little is known about the nature of the venoms, which are associated with the spines of the many stinging fish found in Australian waters...Most of these fish venoms are presumably unstable in heat, and the aim of such treatment is to inactivate the venom present superficially and under the skin."

The text *Venomous and Poisonous Marine Animals, a Medical and Biological Handbook*, ¹⁶ while covering in considerable detail many aspects of catfish envenomation, contains only one direct comment on the possible heat-labile nature of fish venoms:

"Fish venoms are predominantly unstable large proteins (Halstead 1988). As such molecules are dissociated with changes in pH and temperature, hot-water immersion might cause denaturation of the venom in the tissues. However, the return of pain on extraction of the part from hot water casts some doubt on this rationale. The analgesic efficacy of hot-water immersion for venomous fish injuries cannot be disputed and should always be adopted by first-aiders as a first measure for pain relief in venomous fish stings."

In summary, certain facts emerge concerning catfish envenomation. Most (probably all) catfish of the Plotosidae family contain venom apparatus and are a common cause worldwide of fish spine envenomations in humans. No scientific study has ever demonstrated heat-lability of catfish venom (to the best of my knowledge).

Might not the Gate Control and DNIC theories of pain explain the underlying mechanism for fish spine envenomation analgesia by hot water immersion?

Kakigi and Watanabe have shown that interference stimulations using vibration, active and passive movements of the hand or foot, noxious warming by hot water (46°C) and noxious cooling by ice water (0°C) all caused significant reduction in pain perception in normal human volunteers who were experiencing painful stimulation of either ipsilateral or contralateral hand or foot via CO₂ laser.⁵ Specifically, they noted markedly reduced pain amplitude using noxious warming and cooling stimulation applied to the peripheral skin close to and remote from the site where laser stimulation was applied.

They deduced that, since the hot and cold stimuli mainly ascend through the small fibres, this pain relief could be

better accounted for by DNIC theory than Gate Control theory, and they refer to clinical studies indicating that the site responsible for DNIC is the brainstem. Whilst an account of DNIC theory is beyond the scope of this paper, its application in the above study is clearly relevant to the phenomenon of hot water analgesia in fish spine envenomation, particularly as the study used water at 46°C as the noxious stimulus. Further, standardised pain scores three to six minutes after taking the hand from the hot water (after-effect) did not show any significant change from the control session, consistent with my personal experience of rapid return of pain following arm removal from hot water.

Two questions are posed. Has hot water immersion been trialled for above-water envenomations, such as arachnids, hymenoptera and arthropods? Some of these toxins are heat-stable so a demonstrable efficacy would challenge the role of heat-lability as already discussed. Interestingly, although application of hot packs to jellyfish stings has been found to have only mild analgesic effect, immersion of the affected part in hot water has been found to be very effective.⁶

Secondly, would hot-water immersion of the contralateral limb also be effective in marine fish spine envenomations, or even an upper limb immersion where the lower limb is envenomed, or vice versa?

In conclusion, an extensive literature search has failed to find evidence supporting the denaturation theory. Further research is needed to investigate the mechanism(s) underlying pain relief by hot water immersion of the affected limb following fish spine envenomation. The author hypothesises that modern pain theory provides a better explanation than heat denaturation of toxins.

Acknowledgements

I thank Ms Eva McLusky without whose insight this article would not have been written, and Ms Sandra Mangion, formerly Clinical Pharmacist of DATIS (Drugs & Therapeutic Information Service, Repatriation General Hospital, Daw Park, S.A.), for help with the literature search.

References

- 1 Hawdon GM, Winkel KD. Venomous marine creatures. *Aust Fam Physician* 1997; 26: 1369-1374
- 2 Judd M, White J. A South Australian handbook on bites and stings: the animals, the injuries, first aid. Women's and Children's Hospital, The South Australian Museum, The Advertiser 1994 (revised 2000). Adelaide: 41-42
- 3 Fulde G. Emergency medicine quiz. *Current Therapeutics* 2001; 57: 96.
- Power I. How to treat: Management of acute pain Pt
 'Australian Doctor Pull-out Section' Australian Doctor 9 March 2001: III
- 5 Kakigi R, Watanabe S. Pain relief by various kinds of

- interference stimulation applied to the peripheral skin in humans: pain-related brain potentials following CO₂ laser stimulations. *J Peripher Nerv Syst* 1996; 1: 189-198
- 6 Taylor G. Are some jellyfish toxins heat labile? *SPUMS J* 2000; 30: 74-75
- 7 Mann JW 3rd, Werntz JR. Catfish stings to the hand. *J Hand Surg [Am]* 1991; 16: 318-321
- 8 Shiomi K, Takamiya M, Yamanaka H, et al. Hemolytic, lethal and edema-forming activities of the skin secretion from the oriental catfish (Plotosus lineatus). *Toxicon* 1986; 24: 1015-1018
- Scoggin CH. Catfish stings. JAMA 1975; 231: 176-177
- 10 Burnett JW, Calton GJ, Morgan RJ. Catfish poisoning. *Cutis* 1985; 35: 208
- 11 Birkhead WS. The comparative toxicity of stings of the ictalurid catfish genera Ictalurus and Schilbeodes. *Comp Biochem Physiol* 1967; 22: 101-111
- 12 Pacy H. Australian catfish injuries with report of a typical case. *Med J Aust* 1966; 2: 63-65
- 13 Russel FE, Fairchild DM, Michaelson J. Some properties of the venom of the stingray. *Med Arts Sci*

- 1958; 12: 78 (Quoted in Pacy)
- 14 Halstead BW. *Dangerous marine animals*. Cambridge, Maryland: Cornell Maritime Press: 1959. (Quoted in Pacy)
- 15 Isbister GK. Venomous fish stings in tropical northern Australia. *Am J Emerg Med* 2001; 19: 561-565
- Williamson JA, Fenner PJ, Burnett JW, Rifkin JF. Venomous and poisonous marine animals: a medical and biological handbook. Sydney: University Of New South Wales Press; 1966.
- 17 Klasco RK (Ed): POISINDEXTM System. MICROMEDEX, Greenwood Village, Colorado (Healthcare Series Vol 110 expires 12/2001).
- 18 Sutherland SK, Tibballs J. *Australian animal toxins*. 2nd ed. Melbourne: Oxford University Press; 2001.

David Spencer Muirhead, MB BS, Dip RACOG FRACGP is a General Practitioner in full-time practice at Rose Street Clinic, 1 Rose St. Glenelg SA 5045.

Phone: +61-08-8295-2167 *Fax:* +61-08-8376-0906

E-Mail: <dmuirhea@bigpond.net.au>